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(54) Title: MAMMALIAN GAMETE AND EMBRYO CULTURE MEDIA SUPPLEMENT AND METHOD OF USING SAME

(57) Abstract: The present invention provides a supplement and a culture media useful for culturing mammalian gametes and embryonic tissue. The culture media comprises at least one of recombinant human albumin, fermented hyaluronan, and citrate. Because the constituents are produced from non-conventional sources, the culture medium is free from contaminants such as viruses, prions and endotoxins. Additionally, because the medium is completely defined, the medium is not subject to variations which can impair the development of mammalian cells and prevent meaningful comparisons of empirical studies.

WO 01/94552 A2

MAMMALIAN GAMETE AND EMBRYO CULTURE MEDIA SUPPLEMENT AND METHOD OF USING SAME

FIELD OF THE INVENTION

5 The present invention relates to culture media which provide useful environments for cellular development. More particularly this invention relates to defined culture media supplement containing constituents produced from non-traditional sources that, when added to culture media, avoid the problems of prior culture media.

BACKGROUND OF THE INVENTION

10 For years media supplements for culturing mammalian embryonic cells have been derived from animal fluids, and in particular blood serum. While serum based media supplements have been somewhat effective for culturing certain types of cells and tissues, these media supplements have been found to be
15 undesirable. One of the main reasons that these serum based media supplements are unattractive candidates for culturing cells is because of the possibility that the resulting media will be contaminated with impurities, toxins, and infective agents found in the fluid from which the media is derived. Additionally, because the animals
20 from which the blood is collected are different and live in differing environments, the fluids produced by these animals have different components at differing concentrations. One important aspect of serum based medium that has been

-2-

recognized is the requirement of macromolecules in the medium. In an attempt to mimic serum based products, researchers have attempted to add synthetic macromolecules, such as polyvinyl alcohol, to replace the macromolecules in the serum, such as albumin. However, because serum is largely undefined chemically, removing the serum from culture media and attempting to replace only the larger molecules has produced culture media which are less than ideal or ineffective for many purposes because the media are missing essential components.

Accordingly, there is a need for culture media supplements which are as effective as culture media supplements based on blood products while at the same time eliminating potential sources of contamination. Additionally there is a need for standardized culture media.

SUMMARY OF THE INVENTION

The present invention describes a novel, physiologically based completely defined supplement to culture media for mammalian embryonic cells and gametes. This medium supplement may be used with *in vitro* fertilization media, embryo transfer media and embryo cryopreservation media for the mammalian preimplantation embryo, as well as a supplement to media for the development of embryonic stem cells, or any other similar media well known in the art. This supplement contains recombinant human albumin (rHA), fermented hyaluronan (HYN) and/or citrate, and combinations thereof. Addition of this supplement to the culture medium results in equivalent development compared to media supplemented with serum albumin purified from blood.

DETAILED DESCRIPTION OF THE INVENTION

The supplement may comprise recombinant human albumin (rHA) at any appropriate concentrations for the media in which it is to be used. As further

described herein, the use of rHA rather than naturally occurring human serum albumin (HSA) has numerous advantages.

Typically, when the supplement comprises rHA, the supplement comprises between about 0.1 mg/ml to about 20.0 mg/ml of rHA based on the total
5 volume of the medium to which the supplement is added. In one embodiment, about 0.5 mg/ml to about 5.0 mg/ml of rHA is added based on the total volume of the medium.

The efficacy of the supplement can be enhanced by adding fermented hyaluronan (HYN) to the supplement. The addition of the fermented HYN to the
10 media supplement demonstrates positive results.

The phrase "increases the viability of gametes or embryonic cells" as used herein is defined as including the increased development of the embryos to the blastocyst stage in the culture, the ability to hatch from the zona pellucida is increased *in vitro*, and/or an increase in the overall viability of the embryo in
15 embryo cultures when embryos are cultured in a medium containing the supplement of the present invention as compared to being cultured in the same medium without the supplement.

Furthermore, the addition of fermented HYN to the appropriate medium significantly affects the ability of the blastocysts to survive freezing. The
20 use of fermented HYN has several advantages over the use of HYN from a naturally occurring warm blooded vertebrate source such as purified from rooster comb or umbilical cord. By utilizing fermented HYN rather than HYN from a warm blooded vertebrate source, the ability to control the safety and stability of the HYN from different sources and batches is greatly increased.

25 When present, the amount of fermented HYN will generally be at concentrations between about 0.1 mg/ml to about 5.0 mg/ml based on the total volume of the medium. In one embodiment the fermented HYN will be added to the medium at concentrations between about 0.125 mg/ml to about 1.0 mg/ml based on the total volume of the medium.

The supplement can be further augmented by the addition of citrate. In one embodiment, citrate and rHA are both added to the medium supplement, as it has been surprisingly and unexpectedly discovered that the addition of citrate to a medium supplement containing rHA allows the rHA to closely duplicate the

5 properties of HSA or bovine serum albumin (BSA). The addition of the citrate has a further enhancing effect on the development of the cultured cellular material. Any citrate used for media that is well known in the art may be used, including, but not limited to, choline citrate, calcium citrate, citric acid, sodium citrate, and combinations thereof. In one embodiment, sodium citrate is used. The citrate is

10 generally added at concentrations between about 0.1 mM and about 5.0 mM, based on the total volume of the medium. In one embodiment, the citrate is added at concentrations between about 0.1 mM and about 1.0 mM, based on the total volume of the medium.

The medium supplement of the present invention comprises rHA,

15 fermented HYN and/or citrate in any useful combination. In one embodiment, the medium supplement, and the medium to which the medium supplement is added, is free from non-recombinant macromolecules or macromolecules purified from an animal source. In another embodiment, the medium supplement, and the medium to which the medium supplement is added, is free of non-recombinant HSA and/or

20 non-fermented HYN.

This invention is directed to the medium supplement described above, media containing the medium supplement, a method of making the medium supplement, kits containing the medium supplement, and a method of growing embryonic material employing the medium supplement described herein.

25 The present invention includes a method of growing cellular material, in one embodiment embryos, employing the medium supplement described herein such that they can be included in medium at the start of culture, or can be added in a fed-batch or in a continuous manner. Moreover, the components of the medium supplement may be added together, or separately, at different stages of the media

30 production.

This supplement can be added to any appropriate mammalian cellular material culture media well known in the art, including but not limited to, embryo culture media, embryo transfer media and embryo cryopreservation media (to include both freezing and vitrification procedures) for embryos from any mammalian species, and stem cell media. Any media that can support embryo or cell development could be used, which includes, by way of example only, bicarbonate buffered medium, Hepes-buffered or MOPS buffered medium or phosphate buffered saline. Examples of media are G1.2/G2.2, KSOM/KSOMaa, M16, SOF/SOFaa, MTF, P1, Earle's, Hams F-10, M2, Hepes-G1.2, PBS and/or Whitten's. (Gardner and Lane, 1999; Embryo Culture Systems; Handbook of In Vitro Fertilization, CRC Press, Editors: Trounson AO and Gardner DK, 2nd edition, Boca Raton, pp205-264.)

The production of rHA is well known in the art. In one embodiment, rHA is obtained from genetically modified yeast which produce a human albumin protein. One such methodology for the production of rHA from yeast is taught in U.S. Pat. No. 5,612,197.

Fermented hyaluronan (HYN) is obtained by any process well known in the art. One such process is the continuous bacterial fermentation of Streptococcus equi. Hyaluronan is a naturally occurring polymer of repeated disaccharide units of N-acetylglucosamine and D-glucuronic acid. It is widely distributed throughout the body. Typically, the molecular weight of the fermented HYN is 2.3×10^6 kD. The production of HYN from Streptococcus is well known in the art, and any well known process can be used, including those disclosed in Cifonelli JA, Dorfman A. The biosynthesis of hyaluronic acid by group A Streptococcus: The uridine nucleotides of groups A Streptococcus. J. Biological Chemistry 1957; 228: 547-557; Kjems E, Lebech K. Isolation of hyaluronic acid from cultures of streptococci in a chemically defined medium. Acta Path. Microbiol. Scand. 1976 (Sect. B); 84: 162-164; and Markovitz A, et al. The biosynthesis of hyaluronic acid by group A Streptococcus. J. Biological Chemistry 1959; 234(9): 2343-2350.

Other compounds may be added to the medium supplement of the present invention. These include growth factors, as mammalian embryos and cells typically have many receptors for growth factors and the addition of such growth factors may increase the growth rate of the cultured material. Such growth factors include, but are not limited to, Insulin, typically in amounts of 0.1-100 ng/ml; IGF II, typically in amounts of 0.1-100 ng/ml; EGF, typically in amounts of 0.1-100 ng/ml; LIF, typically in amounts of 5-1000 U/ml; PAF, typically in amounts of 0.1-500 μ M; and combinations thereof. All amounts are based on the total volume of the media to which the medium supplement is added.

Medium supplement can be prepared in 2 ways, either as a separate medium supplement that is added to the media after media preparation, or the ingredients of the medium supplement can be added directly to the culture media during media preparation.

By way of example only, the medium supplement may be prepared on its own as follows. Medium supplement rHA may be made into a stock solution by adding either water, saline or medium to make a concentrated stock solution of between 50-500 mg/ml, usually 250 mg/ml. Alternatively, the solution can be obtained as a 250 mg/ml stock solution. Fermented HYN is reconstituted in water, saline or medium, to make a concentrated stock solution of between 10-500 mg/ml, usually 500 mg/ml. This is achieved by adding the water, saline or medium to a flask and adding the desired amount of HYN to the solution. The HYN is then dissolved by rigorous shaking or mixing using a stir bar. For a 500 mg/ml solution, 500 mg of HYN can be added to 1 ml of solution. Citrate is prepared as a stock solution by adding either water, saline or medium to make a concentrated stock solution of between 5-500 mM, usually 500 mM. For a 500 mM stock solution, 0.9605g of citric acid is added to 10 ml of solution. The rHA, fermented HYN and citrate stocks are added together to make a single supplement solution that is added to the final medium as a 100 x times concentrated stock. For 10 ml of medium, 100 μ l of the supplement is added.

rHA can be added directly to the culture medium as either a powder or as a stock solution. The following embodiment is presented by way of example only. The stock solution may added as 100 μ l of 250 mg/ml stock to 9.9 mls of medium. Fermented HYN may be added directly to the culture medium as either a powder or as a stock solution. As a powder, 1.25 mg of HYN may be added to 10 ml of medium. Alternatively, a 125 μ l of a 1% stock solution may be added to 9.9 ml of medium. Citrate may be added directly to the culture medium as either a powder or as a stock solution. As a powder, 9.6 mg may be added to 100 mls of medium, or alternatively, 100 μ l of a 50 mM stock may be added to 9.9 ml of medium.

All patents and publications cited herein are hereby incorporated by reference.

All ranges recited herein include all combinations and subcombinations included within that range limits; therefore, a range from "about 0.1 mg/ml to about 20.0 mg/ml" would include ranges from about 0.125 mg/ml to about 11.5 mg/ml, about 1.0 mg/ml to about 15.0 mg/ml, etc.

The medium supplement of the present invention solves several problems that persist in the art of culturing mammalian cells, tissues, embryos and other related cellular material. One problem with current media is that the cultured mammalian cellular material, particularly embryos, may become contaminated by contaminants such as prions and/or endotoxins found within macromolecular blood products such as human albumin. An advantage of the supplement of the present invention is that it eliminates the potential contamination associated with the use of blood products in media for culturing embryo and other mammalian cellular materials.

Another problem with current media is the difficulty in standardizing such media when using blood products such as serum albumin or other naturally occurring materials. Furthermore, the present invention makes it easier to purify the final cultured product, when the naturally occurring variations and contaminants within the blood products in the media are eliminated.

The present invention eliminates the inherent variation involved when using a biological protein which is often contaminated with other molecules and which differs significantly between different preparations and also between batches within the same preparation. Therefore, the use of recombinant molecules such as rHA enables the formulation of physiological media to be prepared in a standardized fashion. These preparations are endotoxin free, free of prions and are more physiologically compatible than media which are currently used. Current media contain other synthetic macromolecules, such as polyvinyl alcohol or polyvinyl pyrrolidone, which are unable to perform essential physiological functions, such as bind growth factors, and therefore the use of these media result in inferior development of mammalian cellular material.

The invention will be better understood from the Examples which follow. However, one skilled in the art will readily appreciate that the specific compositions, methods and results discussed are merely illustrative of the invention and no limitations on the invention are implied.

EXAMPLES

Example 1

Media G1.2/G2.2 were prepared from concentrated stock solutions as shown below in Table 1. rHA was added as a 250 mg/ml stock solution of 200 μ l to 9.8 mls of media. Initial experiments have investigated replacing albumin purified from blood with the rHA for outbred mouse embryo development in culture. Fertilized eggs were cultured for 4 days in one of 3 different concentrations of rHA. Embryos were cultured at 37°C in 6%CO₂:5%O₂:89%N₂ in an embryo incubation volume of 10 embryos:20 μ l of medium. Embryos were cultured in medium G 1.2 for 48h followed by 48h of culture in medium G 2.2. The negative control treatment was no protein, the positive control treatment was 5 mg/ml HSA (blood product). The results are shown below in Table 2.

Table 1

Stock	Expires	Components	G1.2 (g/L)	G2.2 (g/L)	FG1 (g/L)
A x10 conc	3 months	NaCl	5.26	5.26	5.844
		KCl	0.41	0.41	0.41
		NaH ₂ PO ₄ -H ₂ O	0.035	0.035	0.078
		MgSO ₄ -7H ₂ O	0.246	0.246	0.246
		Na Lactate	1.17	0.66	0.58
		Glucose	0.09	0.568	0.567
		Penicillin	0.06	0.06	0.06
B x10 conc	1 week	NaHCO ₃	2.101	2.101	2.1
		Phenol Red	0.001	0.001	0.001
C x100 conc	2 weeks	Pyruvic Acid	0.0352	0.011	0.0352
D x100 conc	1 month	CaCl ₂ -H ₂ O	0.265	0.265	0.265
G x100 conc	3 months	alanyl-Glutamine	0.108	0.217	-
T x100 conc	3 months	Taurine	0.0125	-	0.0125
ED	1 month	EDTA	0.029	-	-
		NaOH solution	0.4		
N x100 soln		Non-Essential Amino Acids	10 ml	10 ml	10 ml
E x50 soln		Essential Amino Acids	-	20 ml	-
V x100 soln		Vitamins	-	- 10 mls	10 ml

Stock A and B

1. Weigh out individual components into a 100 ml flask.
2. Add 50 ml of H₂O (either Extreme H₂O or Biowittaker).
3. Mix well until all components are dissolved.
4. Add a further 50 ml of H₂O.
5. Mix well.
6. Filter through 0.2 μ m filter.
7. Store at 4 degrees Celsius.

-10-

Stock C - T

1. Weigh component into a 10 ml tube.
2. Add 10 ml of H₂O (either Extreme H₂O or Biowittaker).
3. Mix well until dissolved.
- 5 4. Filter through 0.2 μ m filter.
5. Store at 4 degrees Celsius.

Embryo Culture Media Preparation - Part I**Stock EDTA**

1. Weigh 0.029g of EDTA into a 10 ml tube.
- 10 2. Weigh 0.4g of NaOH into a separate 10 ml tube.
3. Add 10 ml of H₂O (either Extreme H₂O or Biowittaker) to NaOH and mix until dissolved.
4. Add 220 μ l of NaOH solution to EDTA.
5. Mix until dissolved.
- 15 6. Add 9.8 ml of H₂O to the EDTA.
7. Add 90 ml of H₂O to a 100 ml flask.
8. Add 10 ml of EDTA solution to the 90 ml of H₂O.
9. Filter through 0.2 μ m filter.
10. Store at 4 degrees Celsius.

20 **Table 2**

rHA mg/ml	Blastocyst	Hatching	Cell Number	ICM Number	TE Number	% ICM/Total
0	76.7	31.7	60.8 \pm 2.2 ^a	13.8 \pm 0.7 ^a	47.1 \pm 2.0 ^a	23.0 \pm 0.9
1.25	70.7	46.6	72.6 \pm 2.2 ^{bc}	17.7 \pm 0.6 ^b	56.4 \pm 1.9 ^{bc}	24.0 \pm 0.6
2.5	75	39.3	78.1 \pm 2.5 ^b	18.4 \pm 0.5 ^b	58.4 \pm 2.1 ^b	24.2 \pm 0.5
5	76.8	37.5	65.9 \pm 2.7 ^{ac}	16.3 \pm 0.7 ^b	49.6 \pm 2.4 ^{ac}	25.2 \pm 0.7
HSA 5mg/ml	72.6	38.7	74.3 \pm 2.4 ^{bc}	17.2 \pm 0.7 ^b	56.2 \pm 2.0 ^{bc}	23.6 \pm 0.6

* Different superscripts are significantly different, P < 0.05.

rHA was able to replace HSA for embryo development in culture for at least concentrations of 1.25 to 2.5 mg/ml.

Example 2

- 25 Media G1.2/G2.2 were prepared from concentrated stock solutions as taught in Example 1. Fermented HYN was added from a x100 stock solution of 100 μ l to 10mls of media.

Initial experiments investigated replacing albumin purified from blood with HYN for outbred mouse embryo development in culture. Fertilized eggs were cultured for 4 days in one of 4 different concentrations of HYN. Embryos were cultured at 37°C in 6%CO₂:5%O₂:89%N₂ in an embryo incubation volume of 10 embryos:20μl of medium. Embryos were cultured in medium G1.2 for 48h followed by 48h of culture in medium G2.2. The negative control treatment was no protein. The results are shown below in Table 3.

Table 3

HYN (mg/ml)	Blastocyst	Hatching	Cell Number	ICM Number	TE Number	% ICM/Total
0	82.4 ^a	38.3 ^a	67.3±2.8 ^a	16.1±0.7 ^a	50.3±2.3 ^a	24.6±0.8 ^a
0.125	88.6 ^a	57.1b ^c	79.6±1.9 ^{bc}	21.2±0.8 ^{bc}	58.7±1.5 ^{bc}	26.5±0.6 ^{ac}
0.25	94.5 ^a	72.7 ^c	74.9±3.4 ^{bc}	21.8±1.2 ^{bc}	51.9±2.8 ^{ac}	29.7±0.8 ^{bc}
0.5	100 ^a	50 ^b	64.2±1.9 ^{ac}	18.0±0.7 ^{ac}	46.7±2.0 ^a	28.3±1.1 ^{bc}
1	61.8 ^b	23.5 ^a	62.0±2.7 ^{ac}	17.5±0.8 ^{ac}	49.1±2.5 ^a	26.4±0.9 ^{ac}

* Different superscripts are significantly different, P < 0.05.

Fermented HYN at least of concentrations from 0.125 to 0.5 mg/ml stimulated mouse embryo development.

Example 3

Media G1.2/G2.2 were prepared from concentrated stock solutions as taught in Example 1. rHA was added as a 250 mg/ml stock solution of 200μl to 9.8 mls of media, fermented HYN was added from a x100 stock solution of 100μl to 10mls of media. Subsequent experiments investigated replacing albumin purified from blood with rHA together with fermented HYN for outbred mouse embryo development in culture. Fertilized eggs were cultured for 4 days. Embryos were cultured at 37°C in 6%CO₂:5%O₂:89%N₂ in an embryo incubation volume of 10 embryos:20μl of medium. Embryos were cultured in medium G 1.2 for 48h followed by 48h of culture in medium G 2.2. The results are shown below in Table 4.

Table 4

Treatment	Cell Number	ICM Number	TN Number	% ICM/Total
5mg/ml HSA	73.3 \pm 1.7	17.8 \pm 0.6	55.5 \pm 1.3	24.3 \pm 0.5
1.25mg/ml rHA + 0.125mg/ml HYN	71.8 \pm 1.6	18.6 \pm 0.5	53.2 \pm 1.3	26.0 \pm 0.5*

* Significantly different from the HSA (blood product).

Culture with rHA and fermented HYN together significantly increase the development of the inner cell mass cells (ICM). Since ICM development is linearly related to ability to develop into a viable fetus, an increase in %ICM is likely to mean an increase in viability.

Example 4

Media G1.2/G2.2 were prepared from concentrated stock solutions as taught in Example 1. rHA was added as a 250 mg/ml stock solution of 200 μ l to 9.8 mls of media, fermented HYN was added from a x100 stock solution of 100 μ l to 10mls of media. Subsequent experiments investigated replacing albumin purified from blood with rHA together with fermented HYN for outbred mouse embryo development after transfer to recipient mice. Fertilized eggs were cultured for 4 days and then transferred at the blastocyst stage to recipient females. Embryos were cultured at 37°C in 6%CO₂:5%O₂:89%N₂ in an embryo incubation volume of 10 embryos:20 μ l of medium. Embryos were cultured in medium G1.2 for 48h followed by 48h of culture in medium G2.2. The results are shown below in Table 5.

Table 5

	Implantation rate (%)	Fetal development (%)	Fetus / implantation site (%)	Weight (mg)
HSA	63.3	43.3	68.4	208
rHA and HYN	65.0	46.7	71.8	207

Culture with rHA together with fermented HYN resulted in equivalent fetal development to those embryos cultured in medium supplemented with HSA (blood product).

Example 5

5 Media G1.2/G2.2 were prepared from concentrated stock as taught in Example 1. rHA was added as a 250 mg/ml stock solution of 200 μ l to 9.8 mls of media, fermented HYN was added from a x100 stock solution of 100 μ l to 10mls of media and citrate was added from a x100 stock solution of 100 μ l to 10mls of media. Experiments were performed to determine whether the further supplementation of
10 rHA and fermented HYN together with citrate increased mouse embryo development in culture. Embryos were cultured from the fertilized egg for 48h with rHA and HYN in the presence or absence of citrate. Embryos were cultured at 37°C in 6%CO₂:5%O₂:89%N₂ in an embryo incubation volume of 10 embryos:20 μ l of medium. Embryos were cultured in medium G1.2 for 48h. The results are shown
15 below in Table 6.

Table 6

	Day 3 mean cell number	% 8 cell	% compacted
no citrate	6.33 \pm 0.17	69.7	7.3
citrate	7.21 \pm 0.13*	81.1*	12.2*

* Significantly different from medium lacking citrate, P < 0.05.

The addition of citrate to medium containing rHA and fermented HYN resulted in a significant increase in embryo development.

Example 6

20 Media G1.2/G2.2 were prepared from concentrated stock solutions as taught in Example 1. rHA was added as a 250 mg/ml stock solution of 200 μ l to 9.8 mls of media, fermented HYN was added from a x100 stock solution of 100 μ l to

10mls of media and citrate was added from a x100 stock solution of 100 μ l to 10mls of media. Experiments were performed to determine whether the further supplementation of rHA and fermented HYN together with citrate increased mouse embryo

development in culture. Embryos were cultured from the fertilized egg for 48h with

- 5 rHA and HYN in the presence or absence of citrate. Embryos were then transferred to culture medium with or without citrate for a further 48h. Embryos were cultured at 37°C in 6%CO₂:5%O₂:89%N₂ in an embryo incubation volume of 10 embryos:20 μ l of medium. Embryos were cultured in medium G1.2 for 48h followed by 48h of culture in medium G2.2. The results are shown below in Table 7.

10 Table 7

Treatment	Morula/ Blastocyst (%)	Total Blastocyst (%)	Hatching (% of Total)	Cell Number	ICM Number	TE Number	% ICM/ Total
-/-	53.9	45.2	13	100.4	28.9	71.5	28.4
+/-	71.3	67.3	16.8	112.6	34.9	77.7	30.7
-/+	68.5	64.8	23.2	94.8	33.4	61.44	34.5
+/+	72.7	62.7	18.2	100.7	30.9	69.8	30.4

* Different superscripts are significantly different, P < 0.05.

As can be seen by the results in Table 7, the addition of citrate significantly increased embryo development.

Example 7

- 15 Media G1.2/G2.2 were prepared from concentrated stock solutions as taught in Example 1. rHA was added as a 250mg/ml stock solution of 200 μ l to 9.8 mls of media, fermented HYN was added from a x100 stock solution of 100 μ l to 10 mls of media and citrate was added from a x100 stock solution of 100 μ l to 10 mls of media.

- 20 Initial experiments in the cow have investigated replacing albumin purified from blood (Bovine serum albumin, BSA) with either rHA or fermented HYN or rHA together with fermented HYN for the development of fertilized eggs

in culture. Fertilized eggs were cultured for 6 to 7 days. Embryos were cultured at 38.5°C in 6%CO₂:5%O₂:89%N₂ in 500μl of medium. Embryos were cultured in medium G1.2 for 72h followed by 72h of culture in medium G2.2. The results are shown below in Table 8.

5 **Table 8**

Treatment	Number of Embryos	Total Blastocyst Day 6	Total Blastocyst Day 7
BSA	592	30.9 ^a	36.8
rHA	583	22.1 ^b	38.4
HYN	549	16.9 ^b	30.4
rHA + HYN	558	27.8 ^a	39.1

* Different superscripts are significantly different, P < 0.05.

The combination of both rHA with the fermented HYN produced equivalent embryo development in culture of cow embryos as that obtained in the presence of BSA.

10 **Example 8**

Media G1.2/G2.2 were prepared from concentrated stock solutions as taught in Example 1. rHA was added as a 250mg/ml stock solution of 200μl to 9.8 mls of media, fermented HYN was added from a x100 stock solution of 100μl to 10 mls of media and citrate was added from a x100 stock solution of 100μl to 10 mls of media.

Subsequent experiments in the cow have investigated replacing albumin purified from blood (Bovine serum albumin, BSA) with rHA with or without citrate for the development of fertilized eggs in culture. Fertilized eggs were cultured for 6 to 7 days. Embryos were cultured at 38.5°C in 6%CO₂:5%O₂:89%N₂ in 500μl of medium. Embryos were cultured in medium G1.2 for 72h followed by 72h of culture in medium G2.2. The results are shown below in Table 9.

Table 9

Treatment	Total Blastocyst Day 6	Day 6 Blastocyst Cell Number	Day 6 Blastocyst Inner Cell Mass Cell Number
BSA	40.2	143 ± 6 ^a	46.2 ± 1.9 ^a
rHA	36.6	123 ± 7 ^b	37.9 ± 1.9 ^b
rHA + citrate	41.4	146 ± 5 ^a	45.3 ± 1.9 ^a

* Different superscripts are significantly different, P < 0.05.

Supplementing rHA with citrate resulted in equivalent cow embryo development in culture compared to those embryos cultured in the presence of BSA.

5 Example 9

Media G1.2/G2.2 were prepared from concentrated stock solutions as taught in Example 1. rHA was added as a 250 mg/ml stock solution of 200 μ l to 9.8 ml of media, fermented HYN was added from a x100 stock solution of 100 μ l to 10 ml of media and citrate was added from a x100 stock solution to 10 ml of media.

- 10 Subsequent experiments in the cow have investigated addition of fermented HYN to rHA with citrate for the development of fertilized eggs in culture, and subsequent ability to freeze them. Fertilized eggs were cultured for 6 to 7 days. Embryos were cultured at 38.5°C in 6%CO₂:5%O₂:89%N₂ in 500 μ l of medium. Embryos were cultured in medium G1.2 for 72h, followed by 72h of culture in
- 15 medium G2.2. Blastocysts were either stained for cell numbers or frozen and subsequently thawed to assess survival. The results are shown below in Table 10.

Table 10

Treatment	Total Blastocyst Day 7	Day 7 Blastocyst Cell Number	Survival and Re-expansion Following Freezing
BSA	42.3	150 ± 10	38.5 ^a
rHA + citrate	50.0	134 ± 10	57.1 ^b
rHA + citrate + HYN	51.1	159 ± 10	80 ^c

* Different superscripts are significantly different, P < 0.05.

Supplementing medium with rHA, citrate and fermented HYN significantly increased the ability of blastocysts to survive freezing and thawing.

Example 10

Media G1.2/G2.2 were prepared from concentrated stock solutions as taught in Example 1.

This experiment investigated the effects of growing CF1 mouse embryos in culture in the presence of rHA and HYN on the ability of the embryos to survive freezing and thawing. CF1 mouse embryos were cultured to the blastocyst stage and development and ability to survive the freezing procedure was assessed.

Table 11

Treatment	Development to the Blastocyst Stage (%)	Blastocyst Hatching Rates (%)	Re-expansion After Freezing (%)	Hatching After Freezing (%)	Completely Hatched After Freezing
HSA	88.2	49.0	76.1	42.9	28.6
HSA + HYN	81.8	43.2	79.5	45.5	29.6
rHA + citrate	85.0	53.4	77.5	57.5*	40.0*
rHA + citrate + HYN	79.0	51.9	83.8	67.6*	51.3*

* Significantly different from HSA, $P < 0.05$

From these results it can be clearly seen that culture with rHA or rHA with HYN significantly increases blastocyst hatching after thawing compared to blastocysts grown with HSA ($P < 0.05$).

The ability of the blastocysts to outgrow in culture following cryopreservation was also assessed. The outgrowth of both the ICM and TE was scored between 0-3 where 0 represented no outgrowth and 3 represented extensive outgrowth. Outgrowth has been shown to be related to viability (Lane and Gardner, 1997).

Table 12

Treatment	Attachment (%)	Outgrowth of ICM (%)	Outgrowth of TE (%)
HSA	89.5	0.8±0.1	1.8±0.1
HSA + HYN	91.2	2.2±0.1*	1.7±0.1*
rHA + citrate	85.0	1.8±0.1*	1.6±0.1*
rHA + citrate + HYN	86.8	2.1±0.1*	1.9±0.1*

* Significantly different from HSA, $P < 0.05$

As can be seen from Table 12, development of the ICM was increased by culturing the embryos in a medium containing rHA or HYN as compared to embryos cultured in human serum albumin.

Example 11

This example illustrates that a medium containing rHA, HYN and citrate allows for the successful expansion of cryopreserved supernumerary blastocysts.

In this example, donated cryopreserved human pronucleate embryos were thawed and cultured in medium G1.3 for 48 hours followed by culturing in medium G2.3, as taught in United States Patent Application No. 09/201,594 with the following changes. The G1.2-G1.3 media has a MgSO_4 concentration from 1.0 to 1.8 and a CaCl_2 concentration from 1.8 to 1.0. The changes for the G2.2-G2.3 media are the same as the changes to the G1 media, with the addition on the essential amino acids added at half the concentration and nicotinamide, inositol, and folic acid are not present.

Both media were supplemented with 2.5 mg/ml rHA and 0.125 mg/ml HYN. The freezing solution for the embryos was 4.5% glycerol and 0.1M sucrose (10 min.) followed by 9% glycerol and 0.2M sucrose (7 min.). The embryos were placed in a freezing machine at -6°C , seeded and held for 10 minutes, followed by cooling at 0.5°C per minute to -32°C . The embryos were then plunged into liquid nitrogen. Immediately post thaw, the embryos were incubated individually in 500 nl of fresh G 2.3 for 4 hours after which they were placed

individually in 10 microliters of G 2.3 for overnight culture. All incubations took place in 5% O₂:6% CO₂:89% N₂. The 500 nl samples of media were frozen and analyzed using ultramicrofluorescence. The glucose and pyruvate uptake of the thawed embryos was also measured.

5 **Table 13**

Number of Blastocysts	Mean Glucose Uptake (pmol/embryo/h)	Mean Pyruvate Uptake (pmol/embryo/h)	Number of Blastocysts Completely Expanded After 24h	Number of Blastocysts Completely Hatched After 24h
16	40.6	15.2	12 (75%)	5 (31%)

Example 12

The IVF protocol as outlined in Gardner *et al.* 1988 and Schoolcraft 1999 were used in this example.

This example demonstrates the advantages of a medium containing
 10 rHA, HYN and citrate on the development of human embryos.

Table 14

Treatment Group	Number of Patients	Resulting Pregnancies	Implantation Rates
HSA	10	7 (70%)	32.8%
RHA + citrate + HYN	12	9 (66.7%)	31.9%

CLAIMS

What is claimed is:

- 1 1. A mammalian culture medium supplement comprising
2 recombinant human albumin and fermented hyaluronan, wherein the supplement
3 increases the viability of gametes or embryonic cells cultured in a medium
4 containing the supplement.
- 1 2. The supplement according to claim 1 further comprising
2 citrate.
- 1 3. The supplement according to claim 1, wherein the supplement
2 is free from one or more of non-recombinant macromolecules, non-recombinant
3 human albumin, hyaluronan derived from a warm-blooded vertebrate and
4 combinations thereof.
- 1 4. The supplement according to claim 1, wherein the
2 recombinant human albumin is present in a range of about 0.5 mg/ml to about 5.0
3 mg/ml when added to a medium.
- 1 5. The supplement according to claim 1, wherein the fermented
2 hyaluronan is present in a range of about 0.1 mg/ml to about 1.0 mg/ml when added
3 to a medium.
- 1 6. The supplement according to claim 1, wherein the citrate is
2 present in a range of about 0.1 mM to about 1.0 mM when added to a medium.
- 1 7. The supplement according to claim 1 further comprising a
2 medium that can support embryo or cell development, the medium selected from the
3 group consisting of G1.2/G2.2, KSOM/KSOMaa, M16, SOF/SOFaa, MTF, P1,
4 HTF, Earle's, Hams F-10, M2, Hepes-G1.2, Whitten's and PBS.

1 8. The supplement of claim 7 wherein the medium that can
2 support cell development supports embryo development.

1 9. The supplement of claim 7 wherein the medium that can
2 support cell development supports mammalian stem cell development.

1 10. A mammalian culture medium comprising recombinant human
2 albumin and a medium that can support cell development.

1 11. The mammalian culture medium according to claim 10 further
2 comprising citrate.

1 12. The mammalian culture medium according to claim 10 further
2 comprising fermented hyaluronan.

1 13. The mammalian culture medium according to claim 11 further
2 comprising fermented hyaluronan.

1 14. The mammalian culture medium according to claim 12,
2 wherein the fermented hyaluronan is present in a range of about 0.1 mg/ml to about
3 1.0 mg/ml based on the total volume of the mammalian culture medium.

1 15. The mammalian culture medium according to claim 11,
2 wherein the citrate is present in a range of about 0.1 mM to about 1.0 mM based on
3 the total volume of the mammalian culture medium.

1 16. The mammalian culture medium according to claim 10,
2 wherein the recombinant human albumin is present in a range of about 0.5 mg/ml to
3 about 5.0 mg/ml based on the total volume of the mammalian culture medium.

1 17. A mammalian culture medium comprising fermented
2 hyaluronan and a medium that can support cell development.

1 18. The mammalian culture medium according to claim 17 further
2 comprising citrate.

1 19. The mammalian culture medium according to claim 17,
2 wherein the fermented hyaluronan is present in a range of about 0.1 mg/ml to about
3 1.0 mg/ml based on the total volume of the mammalian culture medium.

1 20. The mammalian culture medium according to claim 18,
2 wherein the citrate is present in a range of about 0.1 mM to about 1.0 mM based on
3 the total volume of the mammalian culture medium.

1 21. A method of producing a supplement for a mammalian culture
2 medium comprising adding recombinant human albumin to either water, saline or
3 medium to make a supplement for a mammalian culture medium.

1 22. The method of producing a supplement for a mammalian
2 culture medium of claim 21 further comprising adding fermented hyaluronan.

1 23. The method of producing a supplement for a mammalian
2 culture medium of claim 21 further comprising adding citrate.

1 24. A method of producing a supplement for a mammalian culture
2 medium comprising adding fermented hyaluronan to either water, saline or medium
3 to make a supplement for a mammalian culture medium.

1 25. The method of producing a supplement for a mammalian
2 culture medium of claim 24 further comprising adding citrate.

1 26. A kit for supplementation of mammalian culture medium,
2 comprising:

3 (a) one or more ingredients selected from the group consisting of
4 of mammalian culture medium, recombinant human albumin, fermented hyaluronan,
5 citrate and combinations thereof; and

6 (b) instructions for use of the kit.

1 27. The kit according to claim 26, wherein the kit comprises a
2 mammalian culture medium, wherein the mammalian culture medium is free from
3 one or more of non-recombinant macromolecules, non-recombinant human albumin,
4 and non-fermented hyaluronan.

1 28. The kit according to claim 26, wherein the instructions
2 provide how to make a mammalian culture medium that is free from one or more of
3 non-recombinant macromolecules, non-recombinant human albumin, and non-
4 fermented hyaluronan.

1 29. The kit according to claim 26, wherein the instructions teach
2 how to make a mammalian culture medium comprising one or more of recombinant
3 human albumin in an amount of about 0.5 mg/ml to about 5.0 mg/ml, fermented
4 hyaluronan in an amount of about 0.1 mg/ml to about 1.0 mg/ml, citrate in a
5 concentration of about 0.1 mM to about 1.0 mM, and combinations thereof, based
6 on the total weight of the mammalian culture medium.

1 30. A mammalian culture medium consisting essentially of:
2 (a) a medium that can support mammalian cell development;
3 (b) recombinant human albumin in an amount from about 0.1
4 mg/ml to about 20.0 mg/ml;
5 (c) fermented hyaluronan in an amount from about 0.1 mg/ml
6 to about 5.0 mg/ml; and
7 (d) citrate in a concentration from about 0.1 mM to about
8 5.0 mM.

1 31. The culture medium according to claim 30, wherein the
2 medium that can support embryo or cell development is selected from the group
3 consisting of G1.2/G2.2, KSOM/KSOMaa, M16, SOF/SOFaa, MTF, P1, HTF,
4 Earle's, Hams F-10, M2, Hepes-G1.2, Whitten's and PBS.

1 32. The culture medium according to claim 30, wherein the
2 culture medium is free from one or more of non-recombinant macromolecules, non-
3 recombinant human albumin, hyaluronan derived from a warm-blooded vertebrate
4 and combinations thereof.

1 33. A mammalian culture medium supplement consisting
2 essentially of:

3 (a) recombinant human albumin in an amount from about 0.125
4 mg/ml to about 20.0 mg/ml;

5 (b) fermented hyaluronan in an amount from about 0.1 mg/ml to
6 about 5.0 mg/ml; and

7 (c) citrate in a concentration from about 0.1 mM to about 5.0
8 mM.

1 34. A method of increasing the viability of embryonic cells
2 comprising culturing an embryo in the mammalian culture medium of claim 10,
3 wherein the viability of the embryo is increased.



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Declaration under Rule 4.17:

— *of inventorship (Rule 4.17(iv)) for US only*

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For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: MAMMALIAN GAMETE AND EMBRYO CULTURE MEDIA SUPPLEMENT AND METHOD OF USING SAME

(57) Abstract: The present invention provides a supplement and a culture media useful for culturing mammalian gametes and embryonic tissue. The culture media comprises at least one of recombinant human albumin, fermented hyaluronan, and citrate. Because the constituents are produced from non-conventional sources, the culture medium is free from contaminants such as viruses, prions and endotoxins. Additionally, because the medium is completely defined, the medium is not subject to variations which can impair the development of mammalian cells and prevent meaningful comparisons of empirical studies.

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INTERNATIONAL SEARCH REPORT

Int'l National Application No

/US 01/18766

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C12N5/06

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	EP 0 947 581 A (YOSHITOMI PHARMACEUTICAL) 6 October 1999 (1999-10-06) abstract; claims 1,10 ---	10,16, 21,26-29
X	KEENAN J ET AL: "Recombinant human albumin in cell culture: Evaluation of growth-promoting potential for NRK and SCC-9 cells in vitro." CYTOTECHNOLOGY, vol. 24, no. 3, 1997, pages 243-252, XP001038705 ISSN: 0920-9069 the whole document ---	10,16, 21,26-29
X	WO 00 32140 A (IVF SCIENCES COLORADO INC) 8 June 2000 (2000-06-08) the whole document ---	1-16, 21-23, 26-34
-/--		

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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X document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

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INTERNATIONAL SEARCH REPORT

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PU 01/18766

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P, X	HOOPER K. ET AL: "TOWARD DEFINED PHYSIOLOGICAL EMBRYO CULTURE MEDIA: REPLACEMENT OF BSA WITH RECOMBINANT ALBUMIN" BIOLOGY OF REPRODUCTION, SOCIETY FOR THE STUDY OF REPRODUCTION, CHAMPAIGN, IL, US, vol. 62, no. SUPPL 1, 15 July 2000 (2000-07-15), pages 249-250, XP001038047 ISSN: 0006-3363 abstract	10, 11, 15, 16, 21, 23, 26-29, 34
A	--- GARDNER DAVID K ET AL: "Culture of viable human blastocysts in defined sequential serum-free media." HUMAN REPRODUCTION (OXFORD), vol. 13, no. SUPPL. 3, June 1998 (1998-06), pages 148-159, XP001026692 ISSN: 0268-1161 the whole document -----	1

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US 01/18766

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

Although claim 34 is directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2. ☐ Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

see isa/206

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

International application No.

PCT/US 01/18766

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1-9, 12-14, 22, 26-33 all partially and 10, 11, 15, 16, 21, 23, 34 all completely

Mammalian culture media supplements comprising recombinant human albumin, methods for producing such, kits including such and mammalian culture medium comprising such.

2. Claims: 1-9, 12-14, 22, 26-33 all partially and 17-20, 24, 25 all completely

Mammalian culture media supplements comprising fermented hyaluronan, methods for producing such, kits including such and mammalian culture medium comprising such.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

/US 01/18766

Patent document cited in search report		Publication date	Patent family member(s)	Publication date
EP 0947581	A	06-10-1999	WO 9806822 A	19-02-1998
WO 0032140	A	08-06-2000	AU 2707100 A	19-06-2000
			EP 1146837 A	24-10-2001